Guidelines for the minimum set of photos of accessions submitted for field verification

Developed by the Taxonomy Advisory Group (TAG)

Purpose and technical advice

The purpose of this list is to assist you in taking the most appropriate photos to illustrate the field verification work and eventually, seek the opinion of the Taxonomy Advisory Group regarding the identification of uncertain accessions. You can post the set on the TAG forum: <u>http://tag.inibap.org/</u>. The guidelines are compatible with the requirements of MGIS in order to able to download them directly into the database.

This is a suggested minimum set of photos that can be completed with more photos, if necessary. Sometimes, the development stage of the plant will not allow you to take the full set at the same time. In that case, photograph the characters that you think are most likely to help in the identification. The schemas in this document can help you identify the character. We would appreciate if you could fill out the form at the end of this document (one form per accession).

For the field verification exercise, also take photos of the <u>somaclonal variants</u>, if there are any. Send us the photos regardless of the age of the plant. Sometime the abnormalities are obvious when the plant is young, but sometimes they appear at a later stage, even during the second cycle.

Technical information about the camera used should be given with the photos (see Angela Kepler's guidelines). A "macro" position is advisable to take good close-up photos.

Photos have to come with at least the following information when they are submitted to Bioversity:

- The filename has to include the photo number (1 to 15), the ITC code, the cultivar's name and the part of the plant photographed.
 e.g. "8_ ITC0177_Makara_Bract and flowers_"
- Captions including the location, accession name, accession code, crop cycle and author also has to be provided in an Excel or Word file.

Bring with you the booklet "**Descriptors for bananas**, *Musa* **spp**." **and its colour chart**. For all photos where colour is important (such as photo 7, 8, 10, 13, 14), include the *Musa* colour chart in the photo for comparison purposes. Also bring with you the addendum for East African Highland Bananas if you have to photograph EAHB accessions.

1

Guidelines for the minimum set of photos for accessions submitted for field verification



The appropriate development stage for observation

You will notice that, between the bunch and the male bud, the nodes are arranged on the rachis along three intertwined long spirals (a node is the scar made by a fallen bract).

Choose one of the spirals and count the number of nodes/scars along it. If there are 20 of them, the plant is at the right development stage for photos and description.

On average, bracts fall off at the rate of one a day. Since there are always three spirals, multiplying the number of nodes/scars in a spiral by three gives the time elapsed since flowering. Twenty nodes mean that the plant flowered 60 days before. This is the point after which rapid change no longer occurs.





Guidelines for the minimum set of photos for accessions submitted for field verification

Photo 5 – Neck

- · Vegetative part to be observed on a tall sucker (ideally at the stage of 20 emerged leaves)
- Photograph the zone where the leaf sheaths separate (i.e. the 'neck' of the pseudostem').
- · Choose the best angle to show the canal type and the leaf bases
- Make a close up on the petiole base in order to see the colour line on the petiole margin, the petiole wings and the blotches at the petiole basis.

6.3 - Leaf Petiol Canal



6.3.3 Petiole canal leaf III Leaf III is the third leaf counted from the last leaf (leaf I) produced before bunch emergence. Cut the petiole half way between the pseudostem and the leaf blade and examine the cross section. (See Figs. 4 and 6) 1 Open with margins spreading 2 Wide with erect margins 3 Straight with erect margins 4 Margins curved inward 5 Margins overlapping

Fig. 6 Petiole canal leaf III

Photo 6 – leaf petiol section

Cut a leaf, preferably the 3rd one. Cut the petiole a third of the way down from the blade (or halfway down if the petiole is long). The cut should be done with a sharp knife to make it a straight one. Photograph the petiole section to show if the leaf canal is opened or closed.

Make an imprint of the cross-section (you can cut off the leaf blade to make it easier). Press the surface of the crosssection on a sheet of white paper. Some sap will flow out and make an imprint on the paper. After a short time (15 to 45 seconds are usually sufficient).that imprint will turn brown, at which point it can be scanned. If you cannot scan it, photograph it.

If there is a lot of sap, one can take a set of imprints which have been left to dry for different lengths of time. If there is very little sap, press longer against the paper. If needed, use a pencil to highlight the petiole wings.



4





6

Guidelines for the minimum set of photos for accessions submitted for field verification 🏄

colour of the flesh.

Beware of sunshine and camera flash washing out the

Add the colour chart B on your photo

00

8

Photo 16 (optional): Extra photo of a character that is very specific to the variety

Minimum list of descriptors for field verification

Please, indicate the result of your field observation regarding the accession status:

Flease, II	iuicate the result of your held observation regardin	ig the accession status.
Results:	1.True to type (TTT) 2. True to Sub-group (T (please circle your choice)	TS) 3. Genetic Deviation (GD) 4. Mislabelling (ML)
Fill in the measuren	e hereunder field form with any complementary c nents) and precise the colours observed with charts. T	observation or character that cannot be seen on the photos (e.g. Fhis form will facilitate data entry in MGIS.
Collection:		Planting date:// Photos date://
Curator/Observer:		Original accession in the collection: YES NO
Accession name:		Number of plants: Cycle number:
Botanical classification:		Number of empty nodes (6.4.2):
TC number: Accession code:		Location in the collection:
Origin:		Description recorded in MGIS: YES N0
<i>If anothe</i> 6.1 PL	er value should be recorded, note "N" and specify in Notes.	
		6.4.6 Bunch position
For t	he following descriptors use Colour Chart A	1 Hanging vertically
6.2.1	Pseudostem height [m]	2 Slightly angled
	1 <u><</u> 2 m	3 Hanging at angle 45°
	2 2.1 to 2.9 m	4 Horizontal
	3 <u>></u> 3 m	5 Erect
6.2 PS	SEUDOSTEM/SUCKERS	6.4.7 Bunch shape
6.2.6	Pigmentation of the underlying pseudostem	2 Iruncated cone snape
	1 Pink-purple	3 ASYMMETIC - BUNCH AXIS IS NEARLY STRAIGHT
	2 Red	4 With a curve in the bunch axis
() 5	3 Purple	o spilal 6.4.12 Dechic position
6.3 PE	ETIOLE/MIDRIB/LEAF	0.4.12 Racins position 1 Falling vortically
() 1	Distance at the methols have	2 Δt an angle
0.3.1	Biolones al ine periore base	3 With a curve
	2 Small blotchas	4 Horizontal
	2 Jilidii Diololies 2 Largo blotobos	5 Frect
	5 Large biolories 4 Extensive nigmentation	6.4.13 Rachis appearance
	5 Without nigmentation	1 Bare
633	Petiole canal leaf III	2 Neutral flowers
0.0.0	1 Open with margins spreading	3 Male flowers/bracts above the male bud
	2 Wide with erect margins	4 Neutral/male flowers and presence of withered
	3 Straight with erect margins	bracts
	4 Margins curved inward	6.4.15 Male bud shape
	5 Margins overlapping	1 Like a top
6.3.4	Petiole margins	2 Lanceolate
	1 Winged and undulating	3 Intermediate
	2 Winged and not clasping the pseudostem	4 Ovoid
	3 Winged and clasping the pseudostem	5 Rounded
	4 Not winged and clasping the pseudostem	6.4.16 Male bud size [cm]
	5 Not winged and not clasping the pseudostem	Length and maximum diameter of male bud at harvest
6.3.6	Petiole margin colour	1. $<$ 01 equal to 20 CM 2. 21 to 20 cm
	I Green	2. $21 \text{ to } 30 \text{ cm}$
	2 Pink/purple to red	5. > 01 Equal to 31 UII 65 RDACT
	3 Purple to blue	
6.3.7	Eage of petiole margin	6.5.1 Bract hase shane
	I Colourless (without a colour line along)	u.u.i Diaci Dase silape 1 Small shoulder
	vviin a colour line along	n Smail Shoulden 2 Medium
	0.0.4 Inhorescence / male bud	2 I arre shoulder

6.5.2 Bract apex shape

- 1 Pointed
 - Slightly pointed 2
 - 3 Intermediate
 - 4 Obtuse
 - 5 Obtuse and split

6.5.3 Bract imbrication

- 1 Old bracts overlap at apex of bud
- Young bracts slightly overlap
 Young bracts greatly overlap

6.5.5 Colour of the bract internal face

- 1 Whitish
 - 2 Yellow or green
 - 3 Orange red
 - 4 Red
 - 5 Purple
 - 6 Purple brown
 - 7 Pink-purple
- 6.5.12 Bract behavior before falling
 - 1 Revolute (*rolling*)
 - 2 Not revolute (not rolling)

6.6 MALE FLOWER

For the following descriptors use Chart B when needed

- 6.6.2 Compound tepal basic colour
 - 1 White
 - 2 Cream
 - 3 Yellow
 - 4 Orange
 - 5 Pink/pink-purple

6.6.4 Lobe colour of compound tepal

- 1 Cream 3 Orange
- 2 Yellow 4 Green
- 6.6.13 Anther colour
 - 5 Brown/rusty brown 1 White
 - 2 Cream 6 Pink/pink-purple
 - 3 Yellow 7 Black (anthers aborted)
 - 4 Grey
- 6.6.24 Dominant colour of male flower
 - 1 White
 - 2 Cream
 - 3 Yellow
 - Pink 4
 - 5 Red-purple

6.7 FRUIT

- 7.10 Number of hands: I
- 6.7.2 Number of fruits on second hand
 - 1 <u><</u>12
 - 2 13-16
 - 3 >17
- 6.7.3 Fruit length [cm]
 - 1 <u><</u>15 cm
 - 2 16-20 cm
 - 3 21-25 cm
 - 4 26-30 cm
 - 5 >31 cm

6.7.4 Fruit shape (longitudinal curvature)

- 1 Straight (or slightly curved)
- 2 Straight in the distal part
- 3 Curved (sharp curve)
- 4 Curved in 'S' shape (double curvature)
- 6.7.6 Fruit apex
 - 1 Pointed
 - 2 Lengthily pointed
 - 3 Blunt-tipped
 - 4 Bottle-necked
 - 5 Rounded

6.7.7 Remains of flower relicts at fruit apex

- 1 Without any floral relicts
- 2 Persistent style
- Base of the style prominent 3
- 6.7.8 Fruit pedicel length [mm]
 - 10 mm 1
 - 2 11 to 20 mm
 - 3 21 mm
- 6.7.11 Fusion of pedicels
 - Very partially or no visible sign of fusion 1
 - 2 Partially fused
 - 3 Totally fused

Notes: