Descriptors for Barley

(Hordeum vulgare L.)





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Financial support for the core programme of IPGRI is provided by the Governments of Australia, Austria, Belgium, Canada, China, Denmark, France, Germany, India, Italy, Japan, the Republic of Korea, the Netherlands, Norway, Spain, Sweden, Switzerland, the UK, the USA, and the World Bank

Citation

IPGRI. 1994. Descriptors for barley (Hordeum vulgare L.). International Plant Genetic Resources Institute, Rome, Italy

ISBN 92-9043-222-5

IPGRI Via delle Sette Chiese 142 00145 Rome Italy

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PREFACE

Descriptors for barley (Hordeum vulgare L.) is a revision of the original IPGRI 'Barley Descriptors' (1982). An updated and slightly modified list was prepared by J.D. Franckowiak and T. Konishi, as recommended by the International Workshop on Barley Genetic Resources held 20-21 July, 1991, in conjunction with the Sixth International Barley Genetics Symposium, Helsingborg, Sweden. A draft version prepared in the IPGRI internationally accepted format for descriptor lists was subsequently sent to a number of experts for their comments and amendments. A full list of the names and addresses of those involved is given in 'Contributors'. The descriptors from the 1982 list are cross-referenced with the current list with the descriptor numbers in parentheses beside the current descriptor.

IPGRI encourages the collection of data for descriptors on the first four categories of this list: Passport, Management, Environment and site, and Characterization; and endorses data in these categories as those that should be available for any one accession. However, the number of each of the site and environment descriptor types used will depend on the crop and their importance to the crop's description. Descriptors listed under Evaluation allow for a more detailed description of the accession's characters, but generally require replicated site and time trials.

Although the suggested coding should not be regarded as the definitive scheme, this format has the full backing of IPGRI and is promoted worldwide. This descriptor list is intended to be comprehensive for the descriptors that it contains. This approach assists with the standardization of descriptor definitions. IPGRI does not, however assume that each curator will characterize accessions of their collection utilizing all descriptors given. Those descriptors known to be highly discriminating descriptors are given in bold and marked with and asterisk. Descriptors should be used when they are useful to the curator for the management and maintenance of the collection and/or to the users of the plant genetic resources.

This descriptor list provides an international format and thereby produces a universally understood 'language' for plant genetic resources data. The adoption of this scheme for data encoding, or at least the production of a transformation method to convert other schemes to the IPGRI format, will produce a rapid, reliable, and efficient means for information storage, retrieval, and communication, and will assist with the utilization of germplasm. It is recommended, therefore, that information should be produced by closely following the descriptor list with regard to: ordering and numbering descriptors; using the descriptors specified; and using the descriptor states recommended.

Any suggestions on this descriptor list will be appreciated by IPGRI.

DEFINITIONS AND USE OF THE DESCRIPTORS

IPGRI now uses the following definitions for genetic resources data management:

- Passport descriptors: These provide the basic information used for the general management of the accession (including the registration at the genebank and other identification information) and describe parameters that should be observed when the accession is originally collected
- Management descriptors: These provide the basis for the management of accessions in the genebank and assist with their multiplication and regeneration
- Environment and site descriptors: These describe the environmental and site specific (iii) parameters that are important when characterization and evaluation trials are held. They can be important for the interpretation of the results of these trials. Germplasm collecting site descriptors are also included here
- Characterization descriptors: These enable an easy and quick discrimination between (iv) phenotypes. They are generally highly heritable, can be easily seen by the eye and are equally expressed in all environments. In addition, these may include a limited number of additional traits thought desirable by a consensus of users of the particular crop
- Evaluation descriptors: Many descriptors in this category are susceptible to environmental differences but are generally useful in crop improvement. In addition, others may involve biochemical or molecular characterization. They include yield, agronomic performance, stress susceptibilities and biochemical and cytological traits

Characterization will normally be the responsibility of genebank curators, while evaluation will typically be carried out elsewhere (possibly by a multidisciplinary team of scientists). The evaluation data should be fed back to the genebank which will maintain a data file.

Minimum descriptors highly discriminating are in bold and marked with an asterisk,

The following internationally accepted norms for the scoring, coding, and recording of descriptor states should be followed as indicated below:

- the Système International d'Unités (SI system) is used. The units to be applied are given in square brackets following the descriptor name;
- (b) standard colour charts, e.g., Royal Horticultural Society Colour Chart, Methuen Handbook of Colour, or Munsell Color Chart for Plant Tissues, are strongly recommended for all ungraded colour characters (the precise chart used should be specified in the section where it is used):

- (c) many quantitative characters which are continuously variable are recorded on a 1-9 scale, where:
 - 1 Very low
 - 2 Very low to low
 - 3 Low
 - 4 Low to intermediate
 - 5 Intermediate
 - 6 Intermediate to high
 - 7 High
 - 8 High to very high
 - 9 Very high

is the expression of a character. The authors of this list have sometimes described only a selection of the states, e.g. 3, 5, and 7 for such descriptors. Where this has occurred, the full range of codes is available for use by extension of the codes given or by interpolation between them - e.g. in Section 10 (Biotic stress susceptibility) 1 = very low susceptibility and 9 = very high susceptibility;

(d) (i) when a descriptor is scored using a 1-9 scale, such as in (c), '0' would be scored when the character is not expressed; and (ii) when a descriptor is inapplicable. In the following example, '0' will be recorded if an accession does not form awns:

Lemma awn barbs

- 3 Smooth (few barbs at tip)
- 5 Intermediate (small barbs on upper half)
- 7 Rough
- (e) absence/presence of characters are scored as in the following example:

Presence/absence of terminal leaflet

- 0 Absent 1 (or +) Present
- (f) blanks are used for information not yet available;
- (g) for accessions which are not generally uniform for a descriptor (e.g. mixed collection, genetic segregation), the mean and standard deviation could be reported where the descriptor is continuous, or where the descriptor is discontinuous up to three codes in the order of frequency could be recorded; or other publicized methods can be utilized, such as van Hintum (1993), that clearly states a method for scoring heterogeneous accessions;

(h) dates should be expressed numerically in the format DDMMYYYY, where

DD - 2 digits to represent the day
MM - 2 digits to represent the month
YYYY - 4 digits to represent the year

PASSPORT

1. ACCESSION DESCRIPTORS

1.1 ACCESSION NUMBER (1.1)

This number serves as a unique identifier for accessions and is assigned when an accession is entered into the collection. Once assigned this number should never be reassigned to another accession in the collection. Even if an accession is lost, its assigned number is still not available for re-use. Letters should be used before the number to identify the genebank or national system (e.g. IDG indicates an accession comes from the genebank at Bari, Italy; CGN indicates an accession comes from the genebank at Wageningen, The Netherlands; PI indicates an accession within the USA system)

1.2 DONOR NAME (1.2)

Name of institution or individual responsible for donating the germplasm

1,3 DONOR NUMBER (1.3)

Number assigned to accession by the donor

1.4 OTHER NUMBER(S) ASSOCIATED WITH THE ACCESSION (1.4)

Any other identification number known to exist in other collections for this accession, e.g. USDA Plant Inventory number (not COLLECTING NUMBER, see 2.2). Other numbers can be added as 1.4.3, etc.

- 1.4.1 Other number 1
- 1.4.2 Other number 2

1.5 SCIENTIFIC NAME (1.5)

- 1.5.1 Genus
- 1.5.2 Species
- 1.5.3 Subspecies
- 1.5.4 Botanical variety

1.6	PEDIGREE	(1.6)
	Parentage or nomenclature, and designations assigned to breeders' material	
	1.6.1 Pedigree/mother line	
	1.6.2 <u>Literature citations</u>	
1.7	CULTIVAR NAME	(1.6)
	Either a registered or other formal cultivar designation given to the accession	
	1.7.1 <u>Cultivar name</u>	
	1.7.2 Year of release of the cultivar/year of registration	
	1.7.3 Other designations assigned to breeder's material	
	1.7.4 <u>Literature citations</u>	
1.8	PRINCIPAL ATTRIBUTE	
	The most useful characteristics of the accession	
1.9	ACQUISITION DATE	(1.7)
	Date on which the accession entered the collection (in the format DDMMYY)	Y)
1.10	ACCESSION SIZE	(1.9)
	Approximate number or weight of seeds of an accession in the genebank	
1.11	TYPE OF MAINTENANCE	
	 Vegetative Seed Vegetative and seed Tissue culture 	

2. COLLECTING DESCRIPTORS

2.1 COLLECTING INSTITUTE(S)

(2.2)

Institute(s) and people collecting/sponsoring the sample collection

2.2 COLLECTING NUMBER

(2.1)

Original number assigned by the collector(s) of the sample, normally composed of the name or initials of the collector(s) followed by a number. This item is essential for identifying duplicates held in different collections and should be unique and always accompany subsamples wherever they are sent

2.3 SITE NUMBER

Number assigned to the physical site by the collector

2.4 COLLECTING DATE OF ORIGINAL SAMPLE

(2.3)

(in the format DDMMYYYY)

2.5 COUNTRY OF COLLECTING

(2.4)

Name of the country in which the sample was collected or was bred. Use the three letter abbreviations from the *International Standard (ISO) Codes for the representation of names of countries*, No. 3166, 1988. Copies of these are available from Beuth Verlag GmbH, Burggrafenstrasse 6, D-10772 Berlin 30, Germany; Tel. 30-2601-2320; Fax 30-2601-1231, Tlx. 1-84-273-din-d

2.6 PROVINCE/STATE

(2.5)

Name of the primary administrative subdivision of the country in which the sample was collected

2.7 DEPARTMENT/COUNTY

Name of the secondary administrative subdivision (within a Province/State) of the country in which the sample was collected

2.8 LOCATION OF COLLECTING SITE

(2.6)

Distance in kilometers and direction from the nearest town, village or map grid reference point (e.g. CURITIBA 7S means 7 km south of Curitiba)

2.9 LATITUDE OF COLLECTING SITE (2.7)Degrees and minutes followed by N (North) or S (South) (e.g. 01030S) 2.10 LONGITUDE OF COLLECTING SITE (2.8)Degrees and minutes followed by E (East) or W (West) (e.g. 07625W) 2.11 ELEVATION OF COLLECTING SITE [m] (2.9)Altitude above sea level 2.12 TYPE OF SAMPLE Form of sample collected. If different types of material were collected from the same source, each sample type should be designated with a unique collecting number and a corresponding unique accession number 1 Vegetative 2 Seed 3 Pollen Tissue culture 2.13 **COLLECTING SOURCE** (2.10)1 Wild habitat 2 Farm land 3 Farm store 4 **Backyard** 5 Market 6 Institute 7 Threshing yard 8 Others (specify in the descriptor COLLECTOR'S NOTES, 2.33) 2.14 STATUS OF SAMPLE (2.11)1 Wild 2 Weedy 3 Landrace 4 Advanced cultivar 5 Breeding/research material 6 Population/composite cross 7 Genetic or cytogenetic stock Other (specify in the descriptor COLLECTOR'S NOTES, 2.33)

2.15 NUMBER OF PLANTS SAMPLED

(2.13)

2.16 NUMBER OF SPIKES COLLECTED

2.17 WEIGHT OF SEED COLLECTED [g]

2.18 MAIN COMPANION SPECIES AT COLLECTING SITE

Was (were) companion species collected?

- 0 No
- + Yes

2.19 CROPPING SYSTEM

- 1 Monoculture
- 2 Intercropped (specify crop(s) in the descriptor COLLECTOR'S NOTES, 2.33)

2.20 PLANT POPULATION DENSITY

- 3 Low
- 5 Medium
- 7 High

2.21 GENETIC EROSION

Estimate of the rate at which genetic erosion of the species is occurring in the region of collecting

- 3 Slow
- 5 Intermediate
- 7 Rapid

2.22 LOCAL/VERNACULAR NAME

(2.12)

Name given by farmer to crop and cultivar/landrace/weed. State language and dialect if the ethnic group is not provided

2.23 ETHNIC GROUP

Name of the tribe of the farmer donating the sample or of the people living in the area of collecting

2.24 PRIMARY GRAIN USAGE

- 1 Human food
- 2 Livestock feed
- 3 Alcoholic beverages
- 4 Other (specify in the descriptor COLLECTOR'S NOTES, 2.33)

2.25 PRIMARY FODDER USAGE

- 1 Grazing
- 2 Dried hay
- 3 Straw for bedding
- 4 Fuel
- 5 Green mature or conservation
- 6 Other (specify in the descriptor COLLECTOR'S NOTES, 2.33)

2.26 COLLECTING SOURCE ENVIRONMENT

Descriptors for Collecting Source Environment (2.26.1 - 2.26.26) are given in Section 6. These are numbered in Section 6 in the following manner 6.1.1 - 6.1.26, but should be used for this section. This has been done in order to reduce the repetition of descriptors in Sections 2 and 5

2.27 CULTURAL PRACTICES

2.27.1 Sowing date

(in the format DDMMYYYY)

2.27.2 Harvest date

(in the format DDMMYYYY)

2.27.3 Irrigation

Specify amount, frequency, and method of application

2.28 PHOTOGRAPH

(2.14)

Was a photograph taken of the accession or habitat at the time of collecting? If so, provide an identification number in the descriptor COLLECTOR'S NOTES, 2.33

- 0 No
- + Yes

2.29 HERBARIUM SPECIMEN

Was a herbarium specimen collected? If so, provide an identification number in the descriptor COLLECTOR'S NOTES, 2.33

- 0 No
- + Yes

2.30 ASSOCIATED FLORA

Other dominant crop species, found at and around the collecting site

2.31 PREVAILING STRESSES

Information on associated biotic and abiotic stresses and the accession's reaction. Indicate if disease indexing was done at the time of collecting

2.32 FREQUENCY OF ACCESSION AT COLLECTING SITE

- 1 Rare
- 3 Occasional
- 5 Frequent
- 7 Abundant
- 9 Very abundant

2.33 COLLECTOR'S NOTES

Additional information recorded by the collector or any specific information on any state in any of the above descriptors

MANAGEMENT

3. SEED MANAGEMENT DESCRIPTORS

3.1	ACCESSION NUMBER	(Passport 1.1)
3.2	POPULATION IDENTIFICATION	(Passport 2.2)
	Collecting number, pedigree, cultivar name, etc. depending on the population type	
3.3	STORAGE ADDRESS	
	(Building, room, shelf numbers/location in medium- and/or long-	term storage)
3.4	STORAGE DATE	
	(in the format DDMMYYYY)	•
3.5	GERMINATION AT STORAGE (INITIAL) [%]	
3.6	DATE OF LAST GERMINATION TEST	
	(in the format DDMMYYYY)	
3.7	GERMINATION AT THE LAST TEST [%]	
3.8	DATE OF NEXT TEST	
	Date when the accession should next be tested (estimate) DDMMYYYY)	(in the format
3.9	MOISTURE CONTENT AT HARVEST [%]	
3.10	MOISTURE CONTENT AT STORAGE (INITIAL) [%]	
3.11	AMOUNT OF SEED IN STORAGE(S) [g or number]	(Passport 1.10)
3.12	DUPLICATION AT OTHER LOCATION(S)	(Passport 1.4)

(Passport 1.1)

4. MULTIPLICATION/REGENERATION DESCRIPTORS

ACCESSION NUMBER

4.2 POPULATION IDENTIFICATION (Passport 2.2) Collecting number, pedigree, cultivar name, etc. depending on the population type 4.3 FIELD PLOT NUMBER 4.4 MULTIPLICATION/REGENERATION SITE LOCATION 4.5 COLLABORATOR 4.6 **SOWING DATE** (in the format DDMMYYYY) 4.7 TRANSPLANTING DATE (in the format DDMMYYYY) SOWING DENSITY 4.8 4.9 **CULTURAL PRACTICES**

4.12 SEEDLING VIGOUR

4.10

4.11

4.1

Assessed 18 days after emergence

GERMINATION IN THE FIELD [%]

FERTILIZER APPLICATION

- 4.13 NUMBER OF PLANTS ESTABLISHED
- 4.14 HARVEST DATE

(in the format DDMMYYYY)

For descriptors 4.15-4.17 only applicable for composite crosses

4.15 NUMBER OF PLANTS POLLINATED

4.16 POLLINATION METHOD

100 or more flowers are preferred

- 1 Selfing
- 2 Chain cross
- 3 Pair crossing
- 4 Bulk pollen
- 5 Isolation
- 6 Cluster bagging
- 4.17 NUMBER OF POLLINATED SPIKES REPRESENTED IN STORE(S)
- 4.18 AGRONOMIC EVALUATION
- 4.19 NUMBER OF TIMES ACCESSION REGENERATED (1.10)
- 4.20 PREVIOUS MULTIPLICATION AND/OR REGENERATION
 - 4.20.1 Location
 - 4.20.2 <u>Sowing date</u> (1.8)

(in the format DDMMYYYY)

- 4.20.3 Plot number
- 4.21 NUMBER OF PLANTS USED IN EACH REGENERATION/MULTIPLICATION
- 4.22 SPECIAL MULTIPLICATION REQUIREMENTS
 - 4.22.1 Cytological
 - 4.22.2 Environmental
 - 4.22.3 Isolation
 - 4.22.4 Gene identification

ENVIRONMENT AND SITE

5. SITE DESCRIPTORS

5.1 COUNTRY OF CHARACTERIZATION AND/OR EVALUATION (3.1, 5.1)

(See instructions in COUNTRY OF COLLECTING, 2.5)

5.2 SITE (RESEARCH INSTITUTE) (3.2, 5.2)

5.2.1 Latitude

Degrees and minutes followed by N (North) or S (South) (e.g. 01030S)

5.2.2 Longitude

Degrees and minutes followed by E (East) or W (West) (e.g. 07625W)

5.2.3 Elevation [m]

Altitude above sea level

5.2.4 Name of farm or institute

EVALUATOR'S NAME AND ADDRESS 5.3 (3.3, 5.3)

SOWING DATE 5.4

(3.4, 5.4)

(in the format DDMMYYYY)

HARVEST DATE 5.5

(3.5, 5.5)

(in the format DDMMYYYY)

5.6 STANDARD CULTIVARS USED

> The applied characteristics to be scored require standardization by comparison with recognized cultivars (e.g. Hordeum vulgare cv. Betzes). The standard cultivars used will be constant at a given site or group of sites; however, specific traits may require other standards

5.7 EVALUATION ENVIRONMENT

Environment in which characterization/evaluation was carried out

- 1 Field
- 2 Screenhouse
- 3 Glasshouse
- 4 Laboratory
- 5 Other (specify in the descriptor NOTES, 5.16)

5.8 PERCENTAGE SEED GERMINATION [%]

Specify number of days over which germination is measured

- 5.9 PERCENTAGE FIELD ESTABLISHMENT [%]
- 5.10 NUMBER OF DAYS TO 50% GERMINATION
- 5.11 SOWING SITE IN FIELD

Give block, strip and/or row/plot numbers as applicable

- 5.12 FIELD SPACING
 - 5.12.1 Distance between plants in a row [cm]
 - 5.12.2 <u>Distance between rows</u> [cm]

5.13 ENVIRONMENTAL CHARACTERISTICS OF THE SITE

Descriptors for the Environmental characteristics of site (5.13.1 - 5.13.26) are given in Section 6. These are numbered in Section 6 in the following manner 6.1.1 - 6.1.26, but should be used for this section. This has been done in order to reduce the repetition of descriptors in Sections 2 and 5

5.14 FERTILIZER

(Specify types, doses, frequency of each, and method of application)

5.15 PLANT PROTECTION

(Specify pesticides used, doses, frequency of each, and method of application)

5.16 NOTES

Any other site-specific information

6. ENVIRONMENT DESCRIPTORS

6.1 COLLECTING AND/OR CHARACTERIZATION/EVALUATION SITE ENVIRONMENT

6.1.1 Topography

This refers to the differences in elevation of the land surface on a broad scale

The reference is:

FAO, 1990. In: Guidelines For Soil Profile Description, 3rd Edition (Revised), Rome, p.70

1	Flat	0	-	0.5%	
2	Almost flat	0.6	-	2.9%	
3	Gently undulating	3	-	5.9%	
4	Undulating	6	-	10.9%	
5	Rolling	11	-	15.9%	
6	Hilly	16	-	30%	
7	Steeply dissected	>30%	6, mc	derate range of elevation	
8	Mountainous	>30%, great range of elevation (>300 m)			
9	Other (specify in the appropriate Section's NOTES)				

6.1.2 <u>Higher level landform</u> (General physiographic features)

The landform refers to the shape of the land surface in the area in which the collecting site is located (Adapted from FAO, 1990)

- 1 Plain
- 2 Basin
- 3 Valley
- 4 Plateau
- 5 Upland
- 6 Hill
- 7 Mountain

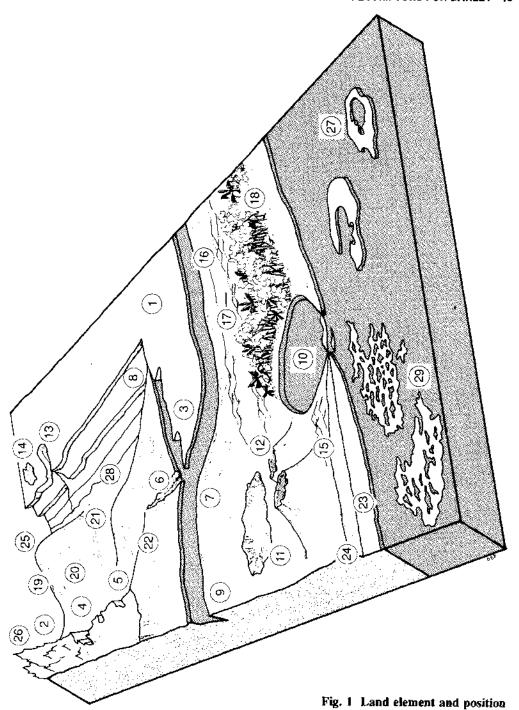
6.1.3 Second level landform (Adapted from FAO, 1990)

1	Alluvial plain	(A plain formed from the deposition of alluvium usually adjacent to a river that periodically overflows (aggraded valley plain, river plain, wash plain, waste plain))
2	Coastal plain	
3	Lacustrine plain	
4	Glacial plain	
5	Peneplain	(Base-leveled plain) (Any land surface changed almost to a plain by subaerial erosion)
6	Pediment	(A piedmont slope formed from a combination of processes which are mainly erosional; the surface is chiefly bare rock but may have a covering veneer of alluvium or gravel (conoplain, piedmont interstream flat))
7	Volcano	<i>"</i>
8	Dunefield	•
9	Delta	
10	Tidal flat	(A marshy, sandy, or muddy nearly horizontal coastal flatland which is alternately covered and exposed as the tide rises and falls)
11	Playa	(A small, generally sandy land area at the mouth of a stream or along the shore of a bay)
12	Cay	(A flat coral island)
13	Other	(Specify in the appropriate Section's NOTES)

6.1.4 Land element and position

Description of the geomorphology of the immediate surroundings of the collecting site (Adapted from FAO, 1990). See Fig. 1

1	Plain level	12 Caldera	23	Beach
2	Escarpment	13 Open depression	24	Beachridge
3	Interfluve	14 Closed depression	25	Rounded summit
4	Valley	15 Dune	26	Summit
5	Valley floor	16 Longitudinal dune	27	Coral atoll
6	Channel	17 Interdunal depression	28	Drainage line (bottom
7	Levee	18 Mangrove		position in flat or
8	Terrace	19 Upper slope		almost flat terrain)
9	Floodplain	20 Mid slope	29	Coral reef
10	Lagoon	21 Lower slope	30	Other (specify in the
11	Pan	22 Ridge		appropriate Section's
		_		NOTES)



6.1.5 Slope [°]

Estimated slope of the collecting site

6.1.6 Slope form

It refers to the general shape of the slope in both the vertical and horizontal directions (FAO, 1990)

- 1 Straight
- 2 Concave
- 3 Convex
- 4 Terraced
- 5 Complex (Irregular)

6.1.7 Slope aspect

The direction that the slope on which the accession was collected faces. Describe the direction with symbols N, S, E, W (e.g. a slope that faces a southwestern direction has an aspect of SW)

6.1.8 Crop agriculture (FAO, 1990)

6.1.8.1 Annual field cropping

- 1 Shifting cultivation
- 2 Fallow system cultivation
- 3 Ley system cultivation
- 4 Rainfed arable cultivation
- 5 Wet rice cultivation
- 6 Irrigated cultivation

6.1.8.2 Perennial field cropping

- 1 Non-irrigated cultivation
- 2 Irrigated cultivation

6.1.8.3 Tree and shrub cropping

- 1 Non-irrigated tree crop cultivation
- 2 Irrigated tree crop cultivation
- 3 Non-irrigated shrub crop cultivation
- 4 Irrigated shrub crop cultivation

6.1.9 Overall vegetation surrounding and at collecting site (FAO, 1990)

1	Grassland	(grasses, subordinate forbs, no woody species)		
2	Forbland	(herbaceous plants predominant)		
3	Forest	(continuous tree layer, crowns overlapping, large number		
		of tree and shrub species in distinct layers)		
4	Woodland	(continuous tree layer, crowns usually not touching,		
		understorey may be present)		
5	Shrubland	(continuous layer of shrubs, crowns touching)		
6	Savanna	(grasses with a discontinuous layer of trees or shrubs)		

6.1.10 Soil parent material (Adapted from FAO, 1990)

Two lists of examples of parent material and rock are given below. The reliability of the geological information and the knowledge of the local lithology will determine whether a general or a specific definition of the parent material can be given. Saprolite is used if the *in situ* weathered material is thoroughly decomposed, clay-rich but still showing rock structure. Alluvial deposits and colluvium derived from a single rock type may be further specified by that rock type

6.1.10.1 Unconsolidated material

- 1 Aeolian deposits (unspecified)
- 2 Aeolian sand
- 3 Littoral deposits
- 4 Lagoonal deposits
- 5 Marine deposits
- 6 Lacustrine deposits
- 7 Fluvial deposits
- 8 Alluvial deposits
- 9 Unconsolidated (unspecified)
- 10 Volcanic ash
- 11 Loess
- 12 Pyroclastic deposits
- 13 Glacial deposits
- 14 Organic deposits
- 15 Colluvial deposits
- 16 In situ weathered
- 17 Saprolite

6.1.10.2 <u>Rock type</u>

1	Acid igneous/	15	Sedimentary rock
	metamorphic rock	16	Limestone
2	Granite	17	Dolomite
3	Gneiss	18	Sandstone
4	Granite/Gneiss	19	Quartzitic sandstone
5	Quartzite	20	Shale
6	Schist	21	Marl
7	Andesite	22	Travertine
8	Diorite	23	Conglomerate
9	Basic igneous/	24	Siltstone
	metamorphic rock	25	Tuff
10	Ultra basic rock	26	Pyroclastic rock
11	Gabbro	27	Evaporite
12	Basalt	28	Gypsum rock

29 Not known

6.1.11 Stoniness/rockiness/hardpan/cementation

13 Dolerite

14 Volcanic rock

- 1 Tillage unaffected
- 2 Tillage affected
- 3 Tillage difficult
- 4 Tillage impossible
- 5 Essentially paved

6.1.12 Soil drainage (Adapted from FAO, 1990)

- 1 Very poorly drained
- 3 Poorly drained
- 5 Moderately drained
- 7 Well drained
- 9 Excessively drained

6.1.13 Flooding (FAO, 1990)

Flooding or temporary inundation is described according to its estimated frequency, duration and depth. Information may be obtained from records of past flooding or from local enquiry. The frequency and duration classes should give an indication of the average occurrence of inundation

6.1.14 Soil depth to groundwater table

(Adapted from FAO, 1990)

The depth to the groundwater table, if present, as well as an estimate of the approximate annual fluctuation, should be given. The maximum rise of the groundwater table can be inferred approximately from changes in profile colour in many, but not all, soils

```
1 (0 - 25 cm)
2 (25.1 - 50 cm)
3 (50.1 - 100 cm)
4 (100.1 - 150 cm)
5 (>150 cm)
```

6.1.15 Quality of the groundwater (FAO, 1990)

- 1 Saline
- 2 Brackish
- 3 Fresh
- 4 Polluted
- 5 Oxygenated
- 6 Stagnating

6.1.16 Soil salinity

- 1 (<160 ppm dissolved salts)
- 2 (161-240 ppm)
- 3 (241-480 ppm)
- 4 (>480 ppm)

6.1.17 Soil moisture (FAO, 1990)

Moisture conditions prevailing in the soil at the time of collecting should be given together with the depth. Attention should be paid to unusual moisture conditions caused by inseasonal weather, prolonged exposure of the profile, flooding, etc.

- 3 Dry
- 5 Slightly moist
- 7 Moist
- 9 Wet

6.1.18 Soil matrix colour (Adapted from FAO, 1990)

The colour of the soil matrix material in the root zone around the accession is recorded in the moist condition (or both dry and moist condition, if possible) using the notation for hue, value and chroma as given in the Munsell Soil Color Charts (Munsell, 1975). If there is no dominant soil matrix colour, the horizon is described as mottled and two or more colours are given and should be registered under uniform conditions. Early morning and late evening readings are not accurate. Provide depth of measurement [cm]. If colour chart is not available, the following categories may be used

1	White	9	Yellow
2	Red	10	Reddish yellow
3	Reddish	11	Greenish, green
4	Yellowish red	12	Grey
5	Brown	13	Greyish
6	Brownish	14	Blue
7	Reddish brown	15	Bluish-black
8	Yellowish brown	16	Black

6.1.19 Soil pH

Actual value of the soil within the following root depths around the accession

6.1.19.1	pH at 10-15 cm
6.1.19.2	pH at 30-60 cm
6.1.19.3	pH at 60-90 cm

6.1.20 Soil organic matter content

- Nil (as on arid zones) 1
- 3 Low (as in long-term cultivation in a tropical setting)
- Medium (as in recently cultivated but not yet much depleted)
- High (as in never cultivated, and in recently cleared from forest)
- 9 Peaty

6.1.21 Rock fragments

Large rock and mineral fragments (>2 mm) are described according to abundance (Adapted from FAO, 1990)

1	(0	-	2%)
2	(2.1	-	5%)
3	(5.1	-	15%)
4	(15.1	-	40%)
5	(40.1	-	80%)
6	(>80%)		

6.1.22 Soil texture classes (Adapted from FAO, 1990)

For convenience in determining the texture classes of the following list and the particle size classes are given for each of the fine earth fraction below. See Fig. 2

1	Clay	12	Coarse sandy loam
2	Loam	13	Loamy sand
3	Clay loam	14	Loamy very fine sand
4	Silt	15	Loamy fine sand
5	Silty clay	16	Loamy coarse sand
6	Silty clay loam	17	Very fine sand
7	Silt loam	18	Fine sand
8	Sandy clay	19	Medium sand
9	Sandy clay loam	20	Coarse sand
10	Sandy loam	21	Sand, unsorted
11	Fine sandy loam	22	Sand, unspecified

1	Clay		<	2 μm
2	Fine silt	3	-	20 μm
3	Coarse silt	21	-	63 µm
4	Very fine sand	64	-	125 µm
5	Fine sand	126	-	200 μm
6	Medium sand	201	-	630 µm
7	Coarse sand	631	-	1250 μm
8	Very coarse sand	1251	-	2000 μm

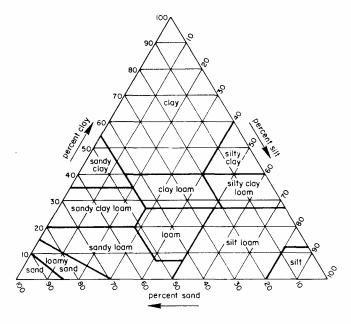


Fig. 2 Soil texture classes

6.1.23 Soil taxonomic classification

As detailed a classification as possible should be given. This may be taken from a soil survey map. State class (e.g., Alfisols, Spodosols, Vertisols etc.)

6.1.24 Water availability

- 1 Rainfed
- 2 Irrigated
- 3 Flooded
- 4 River banks
- 5 Sea coast
- 6 Other (specify in the appropriate Section's NOTES)

6.1.25 Soil fertility

- 3 Low
- 5 Moderate
- 7 High

6.1.26 Climate of collecting site

Should be assessed as close to the collecting as possible

6.1.26.1 <u>Temperature range</u> [°C]

Provide either the diurnal (mean, maximum, minimum) or the seasonal (mean, maximum, minimum)

6.1.26.2 Rainfall range [mm]

Annual average (state number of recorded years)

6.1.26.3 Wind [km s⁻¹]

Annual average (state number of years recorded)

- 6.1.26.3.1 Frequency of typhoons or hurricane force winds
- 6.1.26.3.2 <u>Date of most recent typhoons or hurricane force</u> winds

(in the format DDMMYYYY)

6.1.26.3.3 Annual maximum wind velocity [km s⁻¹]

6.1.26.4	<u>Frost</u>		
	6.1.26	.4.1	Date of most recent frost
			(in the format DDMMYYYY)
	6.1.26	.4.2	Lowest temperature
			Specify seasonal average and minimum survived
	6.1.26	.4.3	<u>Duration of temperature below freezing</u>
6.1.26.5	Relative humidity		
	6.1.26	.5.1	Relative humidity diurnal range [%]
	6.1.26	.5.2	Relative humidity seasonal range [%]
6.1.26.6	Light		
	3 7	Shady Sunny	

6.1.27 Other (specify in the appropriate Section's NOTES)

CHARACTERIZATION

7. PLANT DESCRIPTORS

7.1 VEGETATIVE

* 7.1.1 <u>Growth class</u> (seasonality)

(4.1.1)

- 1 Winter
- 2 Facultative (intermediate)
- 3 Spring

7.1.2 Growth habit

(Subject to environmental factors). See Fig. 3

- 3 Prostrate
- 5 Intermediate
- 7 Erect

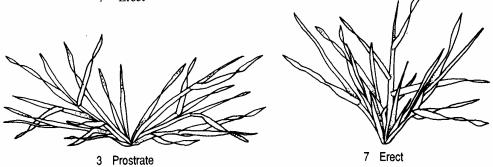


Fig. 3 Growth habit

* 7.1.3 Plant height [cm]

(4.1.2)

At maturity, measured from the ground level to the top of spike excluding awns

* 7.1.4 <u>Stem pigmentation</u> (immature)

- 1 Green
- 2 Purple (basal only)
- 3 Purple (half or more)

7.1.5 Auricle pigmentation

(Very subject to environmental factors)

- 1 Green
- 2 Pale purple
- 3 Purple
- 4 Dark purple

7.2 INFLORESCENCE AND FRUIT

7.2.1 Photoperiod sensitivity

- 1 Very low or no visible sign of sensitivity
- 3 Low sensitivity
- 5 Intermediate
- 7 High sensitivity

7.2.2 Days to flowering

(4.2.1)

From sowing to when 50% of plants have started flowering. However, in dryland areas when planting in dry soils, it is counted from the first day of rainfall or irrigation, which is sufficient for germination. (Highly dependant on temperature-photoperiod interactions)

* 7.2.3 Row number/lateral florets

(4.2.2)

- 1 Two rowed, large or small sterile lateral florets
- 2 Two rowed, deficient
- 3 Irregular, variable lateral florets development
- 4 Six rowed, awnless or awnleted lateral florets
- 5 Six rowed, long awns on lateral florets
- 6 Other (specify in the NOTES descriptor, 7.4)

* 7.2.4 Spike density

(4.2.3)

A subjective observation. See Fig. 4

- 3 Lax
- 5 Intermediate
- 7 Dense

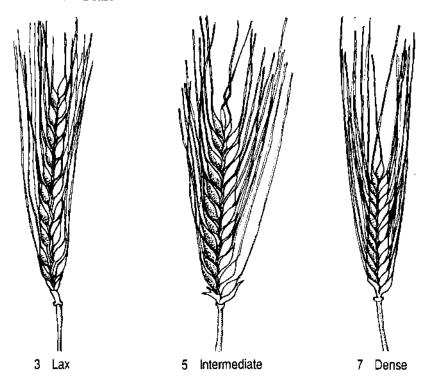


Fig. 4 Spike density

7.2.5 Number of spikelet groups (triplets) per spike

(4.2.4)

An average of five typical spikes selected from a growing accession. (Trait subject to fertility and photoperiod factors)

Lemma awn/hood (4.2.5)7.2.6 See Fig. 5 1 Awnless 2 Awnleted 3 Awned Sessile hoods 4 5 Elevated hoods 1. Awnless 2 Awnleted Sessile hoods 5 Elevated hoods Awned

Fig. 5 Lemma awn/hood

7.2.7 Lemma awn barbs

(4.2.6)

See Fig. 6

- 3 Smooth (few barbs at tip)
- 5 Intermediate (small barbs on upper half)
- 7 Rough

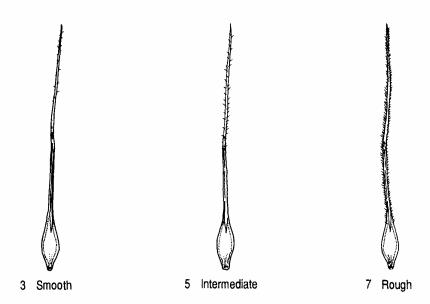


Fig. 6 Lemma awn barbs

7.2.8 Glume and glume awn (outer glumes)

See Fig. 7

- 1 Length of glume and awn shorter than kernel
- 2 Length of glume and awn as long as kernel
- 3 Glume plus awn longer than kernel
- 4 Glume plus awn nearly twice as long as kernel
- 5 Lemma-like
- 6 Other (specify in the descriptor NOTES, 7.4)

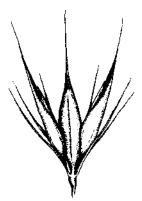
* 7.2.9 Glume colour

- 1 White
- 2 Yellow
- 3 Brown
- 4 Black

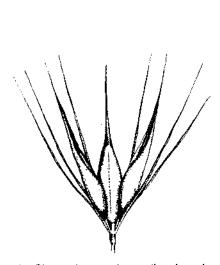
34 DESCRIPTORS FOR BARLEY



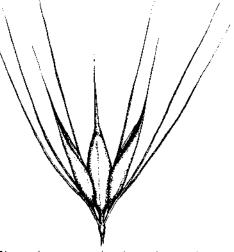
1 Length of glume and awn shorter than kernel



2 Length of glume and awn as long as kernel



3 Glume plus awn longer than kernel



4 Glume plus awn nearly twice as long as kernel

Fig. 7 Glume and glume awn

7.2.10 Lemma type

See Fig. 8

- No lemma teeth
- 2 Lemma teeth (barbs on lateral nerves)
- 3 Lemma hair

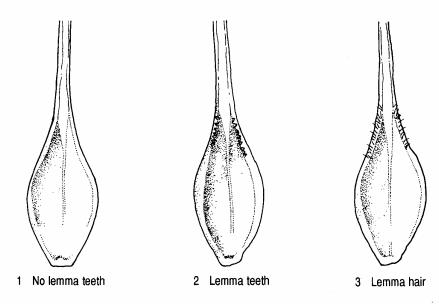


Fig. 8 Lemma type

7.2.11 Awn colour

Recorded on fresh seeds

- Amber/white
- 2 Yellow
- 3 Brown
- 4 Reddish
- 5 Black
- 6 Other (specify in the descriptor NOTES, 7.4)

7.2.12 Length of rachilla hairs

(4.2.7)

See Fig. 9

- Short
- 2 Long

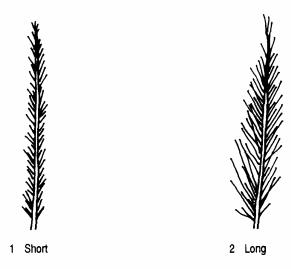


Fig. 9 Length of rachilla hairs

7.3 SEED (Grain)

* 7.3.1 Kernel covering

(4.3.1)

Whether or not the lemma and palea adhere to the caryopsis

- 1 Naked grain
- 2 Semi-covered grain
- 3 Covered grain

7.3.2 Kernel plumpness [%]

Percentage of kernels remaining on a 2.38 mm slotted screen

* 7.3.3 Lemma colour

(4.3.2)

- 1 Amber (=normal)
- 2 Tan/red
- 3 Purple
- 4 Black/grey
- 5 Other (specify in the NOTES descriptor, 7.4)

7.3.4 Grain (pericarp) colour

(4.3.3)

- 1 White
- 2 Tan/red
- 3 Purple
- 4 Black
- 5 Other (specify in the NOTES descriptor, 7.4)

* 7.3.5 <u>Aleurone colour</u>

(Although this trait is difficult to observe, it is used for market type classification in several countries)

- 1 White
- 2 Blue

* 7.3.6 <u>1000-kernel weight</u> [g]

(6.3.1)

(Even though this trait is environment dependent, most growouts will occur in favorable environments). Converted from weighing a lot of 250 kernels or more

7.4 NOTES

Any additional information, especially in the category of 'other' under various descriptors above, may be specified here

EVALUATION

8. PLANT DESCRIPTORS

8.1 SEED (Grain)

8.1.1 Protein content [%] (6.3.2)

Measured as percentage dry weight (seed moisture equal to or less than 12%). Indicating the conversion factors used as either N x 6.25 or N x 5.6

8.1.2 <u>Lysine/protein ratio</u> [%] (6.3.3)

Percentage lysine per unit of protein (absolute)

* 8.1.3 Specific gravity (Test Weight) [kg m⁻³]

Mass of harvested grain of a known volume

- 1 Light (<-1 σ)
- 2 Medium $(+1\sigma)$
- 3 Heavy (>1 σ)

9. ABIOTIC STRESS SUSCEPTIBILITY

Scored under artificial and/or natural conditions, which should be clearly specified. These are coded on a susceptibility scale from 1 to 9 viz.:

- 1 Very low or no visible sign of susceptibility
- 3 Low
- 5 Intermediate
- 7 High
- 9 Very high

9.1 LOW TEMPERATURE

(7.1)

Damage caused by cold to aerial parts of plants (not associated with death of plants in the winter)

9.1.1 Reproductive susceptibility

Measured as reduction in seed set at low temperatures or after frost

9.2	HIGH TEMPERATURE				
9.3	DROUGHT	(7.3)			
9.4	HIGH SOIL MOISTURE	(7.4)			
9.5	WINTER KILL	(7.5)			
	Susceptibility to cold stress measured as a loss of plants in spring (as related to check)				
9.6	SALINITY	(7.6)			
9.7	ACIDITY/ALUMINIUM TOXICITY	(7.7)			
9.8	LODGING				
9.9	AWN RETENTION				
9.10	SHATTERING PRIOR TO HARVEST				
9.11	SPROUTING	(7.8)			

Tendency of grains to sprout in the ear before harvest as a result of late rainfall

10. BIOTIC STRESS SUSCEPTIBILITY

In each case, it is important to state the origin of the infestation or infection, i.e. natural, field inoculation, laboratory. Record such information in the NOTES descriptor, 10.5. Indicate the age of plant when damage is observed.

These are coded on a susceptibility scale from 1 to 9 viz.:

- 1 Very low or no visible sign of susceptibility
- 3 Low
- 5 Intermediate
- 7 High
- 9 Very high

10.1 INSECTS

Causal organism

Disease or common name

10.1.1 Mayetiola destructor

Hessian fly

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	Causal organism	Disease or common name
10.1.2	Oulema melanopus	Cereal leaf beetle
10.1.3	Schizaphis graminum	Greenbug
10.1.4	Diuraphis noxia	Russian wheat aphid
10.1.5	Macrosiphum avenae	English grain aphid
10.1.6	Nematodes (parasitic)	
10.2 FUNG		
10.2.1	Puccinia striiformis f. sp. hordei	Yellow rust
10.2.2	Puccinia graminis f. sp. tritici	Black rust
* 10.2.3	Puccinia hordei	Brown rust, Dwarf leaf rust
10.2.4	Erysiphe graminis f. sp. hordei	Powdery mildew
10.2.5	Rynchosporium secalis	Scald
10.2.6	Pyrenophora graminea	Barley stripe
10.2.7	Pyrenophora teres	Net blotch
10.2.8	Cochliobolus sativus	Spot blotch
10.2.9	Ustilago nuda	Loose smut
10.2.10	<u>Ustilago hordei</u>	Covered smut
10.2.11	Helminthosporium sativum Fusarium spp.	Common root rot
10.2.12	Fusarium graminearum	Scab or head blight
10.2.13	Septoria passerinii	Septoria leaf blotch

		Causal organism	Disease or common name			
	10.2.14	Septoria avenae f. sp. triticea	Speckled leaf blotch			
	10.2.15	Septoria nodorum	Glume blotch			
10.3	BACTE	ERIA				
	10.3.1	Xanthomonas campestris pv. translucens	Bacterial leaf streak			
	10.3.2	Pseudomonas syringae pv. syringae	Bacterial kernel blight			
10.4	VIRUS					
	10.4.1	Barley yellow dwarf virus (BYDV)				
	10.4.2	Barley yellow mosaic virus (BaYMV)				
	10.4.3	Barley stripe mosaic virus (BSMV)	•			
	10.4.4	Barley mild mosaic virus (BaMMV)				
10.5	NOTES					
	Specify	here any additional information or response to o	ther biotic stresses			
11. ALLOENZYME COMPOSITION AND ZYMOTYPE						
11.1	11.1 BIOCHEMICAL MARKERS					
11.1	Diocii	_				

11.1.1 Isozyme

Gene symbols based on nomenclature recommended by the International Barley Genetic Symposium

11.1.2 Other biochemical markers (specify)

11.2 MOLECULAR MARKERS

- 11.2.1 Restriction fragment length polymorphism (RFLP)
- 11.2.2 Random amplified polymorphic DNA (RAPD)

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- 11.2.3 Specific amplicon polymorphism (SAP)
- 11.2.4 Other molecular markers (specify)

12. CYTOLOGICAL CHARACTERS AND IDENTIFIED GENES

12.1 GENETIC STOCK

12.1.1 Marker allele

Specific gene present following the system of nomenclature used in the *Barley Genetic Newsletter*

12.1.2 Mutant stock

Specific trait present based on nomenclature used in the Barley Genetic Newsletter

- 12.1.3 Near-isogenic line
- 12.1.4 Other (specific cytogenetic stocks in descriptor 12.4)

12.2 OTHER GENES PRESENT

Gene symbols of other morphological markers present

12.3 ANEUPLOID OR POLYPLOID STOCK

12.4 STRUCTURAL REARRANGEMENT

- 12.4.1 Translocation
- 12.4.2 Inversion
- 12.4.3 <u>Duplication</u>
- 12.4.4 Deletion
- 12.4.5 Other structural change (specify)

12.5 COMPOSITE CROSS

REFERENCES

- FAO (1990). In: Guidelines For Soil Profile Description, 3rd Edition (Revised). Food and Agriculture Organization of the United Nations, International Soil Reference Information Centre. Land and Water Development Division. Rome, p.70
- Royal Horticultural Society, 1966, c. 1986. R.H.S. colour chart, [ed. 1, 2]. Royal Horticultural Society, London.
- van Hintum, Th. J.L. (1993). In: A computer compatible system for scoring heterogeneous populations. Genetic Resources and Crop Evolution 40: 133-136.

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ACKNOWLEDGEMENTS

IPGRI wishes to place on record their sincere thanks to the numerous barley workers around the world who have contributed to the development of the Descriptors for barley directly or indirectly.

Ms. Adriana Alercia prepared the text for publication. Illustrations were drawn by Mrs. Pina di Pilla. Mr. Paul Stapleton managed the production of the publication. Scientific direction was provided by Dr. Mark Perry.